

Package ‘yarn’

November 28, 2024

Title YARN: Robust Multi-Condition RNA-Seq Preprocessing and Normalization

Version 1.33.0

Description Expedite large RNA-Seq analyses using a combination of previously developed tools. YARN is meant to make it easier for the user in performing basic mis-annotation quality control, filtering, and condition-aware normalization. YARN leverages many Bioconductor tools and statistical techniques to account for the large heterogeneity and sparsity found in very large RNA-seq experiments.

Depends Biobase

License Artistic-2.0

Encoding UTF-8

LazyData true

RoxygenNote 6.1.1

Suggests knitr, rmarkdown, testthat (>= 0.8)

Imports biomaRt, downloader, edgeR, gplots, graphics, limma, matrixStats, preprocessCore, readr, RColorBrewer, stats, quantro

VignetteBuilder knitr

biocViews Software, QualityControl, GeneExpression, Sequencing, Preprocessing, Normalization, Annotation, Visualization, Clustering

git_url <https://git.bioconductor.org/packages/yarn>

git_branch devel

git_last_commit 7e48450

git_last_commit_date 2024-10-29

Repository Bioconductor 3.21

Date/Publication 2024-11-27

Author Joseph N Paulson [aut, cre],
 Cho-Yi Chen [aut],
 Camila Lopes-Ramos [aut],
 Marieke Kuijjer [aut],
 John Platig [aut],
 Abhijeet Sonawane [aut],
 Maud Fagny [aut],
 Kimberly Glass [aut],
 John Quackenbush [aut]

Maintainer Joseph N Paulson <paulson.joseph@gene.com>

Contents

annotateFromBiomart	2
bladder	3
checkMisAnnotation	4
checkTissuesToMerge	5
downloadGTEx	6
extractMatrix	6
filterGenes	7
filterLowGenes	8
filterMissingGenes	8
filterSamples	9
normalizeTissueAware	10
plotCMDS	11
plotDensity	12
plotHeatmap	12
qsmooth	13
qstats	14
skin	15
Index	16

annotateFromBiomart *Annotate your Expression Set with biomaRt*

Description

Annotate your Expression Set with biomaRt

Usage

```
annotateFromBiomart(obj, genes = featureNames(obj),
  filters = "ensembl_gene_id", attributes = c("ensembl_gene_id",
  "hgnc_symbol", "chromosome_name", "start_position", "end_position"),
  biomaRt = "ensembl", dataset = "hsapiens_gene_ensembl", ...)
```

Arguments

obj	ExpressionSet object.
genes	Genes or rownames of the ExpressionSet.
filters	getBM filter value, see getBM help file.
attributes	getBM attributes value, see getBM help file.
biomart	BioMart database name you want to connect to. Possible database names can be retrieved with the function listMarts.
dataset	Dataset you want to use. To see the different datasets available within a biomart you can e.g. do: mart = useMart('ensembl'), followed by listDatasets(mart).
...	Values for useMart, see useMart help file.

Value

ExpressionSet object with a fuller featureData.

Examples

```
data(skin)
# subsetting and changing column name just for a silly example
skin <- skin[1:10,]
colnames(fData(skin)) = paste("names",1:6)
biomart<-"ENSEMBL_MART_ENSEMBL";
genes <- sapply(strsplit(rownames(skin),split="\\."),function(i)i[1])
newskin <-annotateFromBiomart(skin,genes=genes,biomart=biomart)
head(fData(newskin)[,7:11])
```

bladder

Bladder RNA-seq data from the GTEx consortium

Description

Bladder RNA-seq data from the GTEx consortium. V6 release.

Usage

```
data(bladder)
```

Format

An object of class "ExpressionSet"; see [ExpressionSet](#).

Value

ExpressionSet object

Source

GTEEx Portal

References

GTEEx Consortium, 2015. The Genotype-Tissue Expression (GTEEx) pilot analysis: Multitissue gene regulation in humans. *Science*, 348(6235), pp.648-660. ([PubMed](#))

Examples

```
data(bladder);
checkMissAnnotation(bladder);
```

checkMisAnnotation	<i>Check for wrong annotation of a sample using classical MDS and control genes.</i>
--------------------	--

Description

Check for wrong annotation of a sample using classical MDS and control genes.

Usage

```
checkMisAnnotation(obj, phenotype, controlGenes = "all",
  columnID = "chromosome_name", plotFlag = TRUE,
  legendPosition = NULL, ...)
```

Arguments

obj	ExpressionSet object.
phenotype	phenotype column name in the phenoData slot to check.
controlGenes	Name of controlGenes, ie. 'Y' chromosome. Can specify 'all'.
columnID	Column name where controlGenes is defined in the featureData slot if other than 'all'.
plotFlag	TRUE/FALSE Whether to plot or not
legendPosition	Location for the legend.
...	Extra parameters for plotCMDS function.

Value

Plots a classical multi-dimensional scaling of the 'controlGenes'. Optionally returns co-ordinates.

Examples

```
data(bladder)
checkMisAnnotation(bladder, 'GENDER', controlGenes='Y', legendPosition='topleft')
```

checkTissuesToMerge *Check tissues to merge based on gene expression profile*

Description

Check tissues to merge based on gene expression profile

Usage

```
checkTissuesToMerge(obj, majorGroups, minorGroups, filterFun = NULL,  
  plotFlag = TRUE, ...)
```

Arguments

obj	ExpressionSet object.
majorGroups	Column name in the phenoData slot that describes the general body region or site of the sample.
minorGroups	Column name in the phenoData slot that describes the specific body region or site of the sample.
filterFun	Filter group specific genes that might disrupt PCoA analysis.
plotFlag	TRUE/FALSE whether to plot or not
...	Parameters that can go to checkMisAnnotation

Value

CMDS Plots of the majorGroupss colored by the minorGroupss. Optional matrix of CMDS loadings for each comparison.

See Also

checkTissuesToMerge

Examples

```
data(skin)  
checkTissuesToMerge(skin, 'SMTS', 'SMTSD')
```

downloadGTEX	<i>Download GTEX files and turn them into ExpressionSet object</i>
--------------	--

Description

Downloads the V6 GTEX release and turns it into an ExpressionSet object.

Usage

```
downloadGTEX(type = "genes", file = NULL, ...)
```

Arguments

type	Type of counts to download - default genes.
file	File path and name to automatically save the downloaded GTEX expression set. Saves as a RDS file.
...	Does nothing currently.

Value

Organized ExpressionSet set.

Examples

```
# obj <- downloadGTEX(type='genes',file='~/Desktop/gtex.rds')
```

extractMatrix	<i>Extract the appropriate matrix</i>
---------------	---------------------------------------

Description

This returns the raw counts, log₂-transformed raw counts, or normalized expression. If normalized = TRUE then the log parameter is ignored.

Usage

```
extractMatrix(obj, normalized = FALSE, log = TRUE)
```

Arguments

obj	ExpressionSet object or objrix.
normalized	TRUE / FALSE, use the normalized matrix or raw counts
log	TRUE/FALSE log ₂ -transform.

Value

matrix

Examples

```
data(skin)
head(yarn:::extractMatrix(skin,normalized=FALSE,log=TRUE))
head(yarn:::extractMatrix(skin,normalized=FALSE,log=FALSE))
```

filterGenes

Filter specific genes

Description

The main use case for this function is the removal of sex-chromosome genes. Alternatively, filter genes that are not protein-coding.

Usage

```
filterGenes(obj, labels = c("X", "Y", "MT"),
            featureName = "chromosome_name", keepOnly = FALSE)
```

Arguments

obj	ExpressionSet object.
labels	Labels of genes to filter or keep, eg. X, Y, and MT
featureName	FeatureData column name, eg. chr
keepOnly	Filter or keep only the genes with those labels

Value

Filtered ExpressionSet object

Examples

```
data(skin)
filterGenes(skin,labels = c('X','Y','MT'),featureName='chromosome_name')
filterGenes(skin,labels = 'protein_coding',featureName='gene_biotype',keepOnly=TRUE)
```

filterLowGenes	<i>Filter genes that have less than a minimum threshold CPM for a given group/tissue</i>
----------------	--

Description

Filter genes that have less than a minimum threshold CPM for a given group/tissue

Usage

```
filterLowGenes(obj, groups, threshold = 1, minSamples = NULL, ...)
```

Arguments

obj	ExpressionSet object.
groups	Vector of labels for each sample or a column name of the phenoData slot. for the ids to filter. Default is the column names.
threshold	The minimum threshold for calling presence of a gene in a sample.
minSamples	Minimum number of samples - defaults to half the minimum group size.
...	Options for cpm .

Value

Filtered ExpressionSet object

See Also

[cpm](#) function defined in the edgeR package.

Examples

```
data(skin)
filterLowGenes(skin, 'SMTSD')
```

filterMissingGenes	<i>Filter genes not expressed in any sample</i>
--------------------	---

Description

The main use case for this function is the removal of missing genes.

Usage

```
filterMissingGenes(obj, threshold = 0)
```


Arguments

obj ExpressionSet object.
 threshold Minimum sum of gene counts across samples – defaults to zero.

Value

Filtered ExpressionSet object

Examples

```
data(skin)
filterMissingGenes(skin)
```

filterSamples	<i>Filter samples</i>
---------------	-----------------------

Description

Filter samples

Usage

```
filterSamples(obj, ids, groups = colnames(obj), keepOnly = FALSE)
```

Arguments

obj ExpressionSet object.
 ids Names found within the groups labels corresponding to samples to be removed
 groups Vector of labels for each sample or a column name of the phenoData slot for the
 ids to filter. Default is the column names.
 keepOnly Filter or keep only the samples with those labels.

Value

Filtered ExpressionSet object

Examples

```
data(skin)
filterSamples(skin,ids = "Skin - Not Sun Exposed (Suprapubic)",groups="SMTSD")
filterSamples(skin,ids=c("GTEX-OHPL-0008-SM-4E3I9","GTEX-145MN-1526-SM-5SI9T"))
```

normalizeTissueAware *Normalize in a tissue aware context*

Description

This function provides a wrapper to various normalization methods developed. Currently it only wraps qsmooth and quantile normalization returning a log-transformed normalized matrix. qsmooth is a normalization approach that normalizes samples in a condition aware manner.

Usage

```
normalizeTissueAware(obj, groups, normalizationMethod = c("qsmooth",  
  "quantile"), ...)
```

Arguments

obj	ExpressionSet object
groups	Vector of labels for each sample or a column name of the phenoData slot for the ids to filter. Default is the column names
normalizationMethod	Choice of 'qsmooth' or 'quantile'
...	Options for qsmooth function or normalizeQuantiles

Value

ExpressionSet object with an assayData called normalizedMatrix

Source

The function qsmooth comes from the qsmooth packages currently available on github under user 'kokrah'.

Examples

```
data(skin)  
normalizeTissueAware(skin, "SMTSD")
```

`plotCMDS`*Plot classical MDS of dataset*

Description

This function plots the MDS coordinates for the "n" features of interest. Potentially uncovering batch effects or feature relationships.

Usage

```
plotCMDS(obj, comp = 1:2, normalized = FALSE, distFun = dist,  
  distMethod = "euclidian", n = NULL, samples = TRUE, log = TRUE,  
  plotFlag = TRUE, ...)
```

Arguments

<code>obj</code>	ExpressionSet object or objrix.
<code>comp</code>	Which components to display.
<code>normalized</code>	TRUE / FALSE, use the normalized matrix or raw counts.
<code>distFun</code>	Distance function, default is <code>dist</code> .
<code>distMethod</code>	The distance measure to be used. This must be one of "euclidean", "maximum", "manhattan", "canberra", "binary" or "minkowski". Any unambiguous substring can be given.
<code>n</code>	Number of features to make use of in calculating your distances.
<code>samples</code>	Perform on samples or genes.
<code>log</code>	TRUE/FALSE log2-transform raw counts.
<code>plotFlag</code>	TRUE/FALSE whether to plot or not.
<code>...</code>	Additional plot arguments.

Value

coordinates

Examples

```
data(skin)  
res <- plotCMDS(skin, pch=21, bg=factor(pData(skin)$SMTSD))  
  
# library(calibrate)  
# textxy(X=res[,1], Y=res[,2], labs=rownames(res))
```

plotDensity *Density plots of columns in a matrix*

Description

Plots the density of the columns of a matrix. Wrapper for [matdensity](#).

Usage

```
plotDensity(obj, groups = NULL, normalized = FALSE, legendPos = NULL,
  ...)
```

Arguments

obj	ExpressionSet object
groups	Vector of labels for each sample or a column name of the phenoData slot for the ids to filter. Default is the column names.
normalized	TRUE / FALSE, use the normalized matrix or log2-transformed raw counts
legendPos	Legend title position. If null, does not create legend by default.
...	Extra parameters for matdensity .

Value

A density plot for each column in the ExpressionSet object colored by groups

Examples

```
data(skin)
filtData <- filterLowGenes(skin, "SMTSD")
plotDensity(filtData, groups="SMTSD", legendPos="topleft")
# to remove the legend
plotDensity(filtData, groups="SMTSD")
```

plotHeatmap *Plot heatmap of most variable genes*

Description

This function plots a heatmap of the gene expressions for the "n" features of interest.

Usage

```
plotHeatmap(obj, n = NULL, fun = stats::sd, normalized = TRUE,
  log = TRUE, ...)
```

Arguments

obj	ExpressionSet object or objrix.
n	Number of features to make use of in plotting heatmap.
fun	Function to sort genes by, default sd .
normalized	TRUE / FALSE, use the normalized matrix or raw counts.
log	TRUE/FALSE log2-transform raw counts.
...	Additional plot arguments for heatmap.2 .

Value

coordinates

Examples

```
data(skin)
tissues <- pData(skin)$SMTSD
plotHeatmap(skin, normalized=FALSE, log=TRUE, trace="none", n=10)
# Even prettier

# library(RColorBrewer)
data(skin)
tissues <- pData(skin)$SMTSD
heatmapColColors <- brewer.pal(12, "Set3")[as.integer(factor(tissues))]
heatmapCols <- colorRampPalette(brewer.pal(9, "RdBu"))(50)
plotHeatmap(skin, normalized=FALSE, log=TRUE, trace="none", n=10,
  col = heatmapCols, ColSideColors = heatmapColColors, cexRow = 0.6, cexCol = 0.6)
```

qsmooth

Quantile shrinkage normalization

Description

This function was modified from github user kokrah.

Usage

```
qsmooth(obj, groups, norm.factors = NULL, plot = FALSE,
  window = 0.05, log = TRUE)
```

Arguments

obj	for counts use $\log_2(\text{raw counts} + 1)$, for MA use $\log_2(\text{raw intensities})$
groups	groups to which samples belong (character vector)
norm.factors	scaling normalization factors
plot	plot weights? (default=FALSE)

window window size for running median (a fraction of the number of rows of exprs)
log Whether or not the data should be log transformed before normalization, TRUE
 = YES.

Value

Normalized expression

Source

[Kwame Okrah's qsmooth R package](#)

Examples

```
data(skin)
head(yarn:::qsmooth(skin, groups=pData(skin)$SMTSD))
```

qstats

Compute quantile statistics

Description

This function was directly borrowed from github user kokrah.

Usage

```
qstats(exprs, groups, window)
```

Arguments

exprs for counts use $\log_2(\text{raw counts} + 1)$, for MA use $\log_2(\text{raw intensities})$
groups groups to which samples belong (character vector)
window window size for running median as a fraction on the number of rows of exprs

Value

list of statistics

Source

[Kwame Okrah's qsmooth R package](#) Compute quantile statistics

skin

Skin RNA-seq data from the GTEx consortium

Description

Skin RNA-seq data from the GTEx consortium. V6 release. Random selection of 20 skin samples. 13 of the samples are fibroblast cells, 5 Skin sun exposed, 2 sun unexposed.

Usage

```
data(skin)
```

Format

An object of class "ExpressionSet"; see [ExpressionSet](#).

Value

ExpressionSet object

Source

GTEx Portal

References

GTEx Consortium, 2015. The Genotype-Tissue Expression (GTEx) pilot analysis: Multitissue gene regulation in humans. *Science*, 348(6235), pp.648-660. ([PubMed](#))

Examples

```
data(skin);  
checkMissAnnotation(skin, "GENDER");
```

Index

* datasets

bladder, [3](#)

skin, [15](#)

annotateFromBiomart, [2](#)

bladder, [3](#)

checkMisAnnotation, [4](#), [5](#)

checkTissuesToMerge, [5](#)

cpm, [8](#)

downloadGTEx, [6](#)

ExpressionSet, [3](#), [15](#)

extractMatrix, [6](#)

filterGenes, [7](#)

filterLowGenes, [8](#)

filterMissingGenes, [8](#)

filterSamples, [9](#)

heatmap.2, [13](#)

matdensity, [12](#)

normalizeQuantiles, [10](#)

normalizeTissueAware, [10](#)

plotCMDS, [4](#), [11](#)

plotDensity, [12](#)

plotHeatmap, [12](#)

qsmooth, [10](#), [13](#)

qstats, [14](#)

sd, [13](#)

skin, [15](#)